

Sydney E. Everhart

Assistant Professor
Department Plant Pathology
University of Nebraska-Lincoln

406 Plant Sciences Hall
Lincoln, NE 68583-0722
Office: (402) 472-2879
E-mail: everhart@unl.edu
Website: everhart.unl.edu

Education: Ph.D. 2012 University of Georgia (UGA) in Plant Pathology, M.S. 2007 University of Central Missouri (UCM) in Biology, B.S. 2005 University of Iowa in Biology

Research Positions:

- 2016 – Adjunct Assistant Professor, University of Clemson, Clemson, SC
2014 – Assistant Professor/Quantitative Ecologist, Department of Plant Pathology, University of Nebraska (UNL), Lincoln, NE
2012 – 2014 USDA-AFRI-NIFA Postdoctoral Fellow, Department of Botany and Plant Pathology, Oregon State University, Corvallis, OR
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Areas of Expertise and Research Focus: Spatiotemporal epidemiology, fungicide resistance, fungal genetics, whole-genome sequencing, meta-barcode sequencing, development of genetic markers, multivariate analysis of community and population genetic data in R. The focus of research in my lab is on economically important crops (primarily legumes) and pathogens where host resistance is limited.

Highlights:

- Awards and Honors: 25
- Peer-reviewed Publications (since 2008): 27
- First or Corresponding Author on Peer-Reviewed Manuscripts: 15
- Books, Chapters, Proceedings, and R Packages: 6
- Poster and Oral Presentations by a Member of Everhart Lab since 2014: 28
- Students and Postdoctoral Scholars Supervised: 20
- Taught Intro to R workshop to >200 people at local, regional, and international meetings

Manuscript Recognitions:

- Kamvar *et al.* 2017 rated “**Top 5 Most Viewed Articles of 2017**” in *PeerJ*’s sections on Agriculture Science, Genetics, and Mycology
- Grünwald *et al.* 2017 selected for **Sept. 2017 Phytopathology Cover Image**
- Dugan and Everhart 2017 selected as **Feb. 2017 Plant Health Progress Editor’s Pick**

Awards and Honors (25 total):

- 2016:** Schroth Faces of the Future Award, American Phytopathological Society (APS);
Faculty Travel Award UNL
- 2015:** Faculty Travel Award, UNL
- 2012:** Oömycete Bioinformatics Workshop Travel Award
Oömycete Molecular Genetics Meeting Travel Award
Postdoctoral Fellowship, USDA-AFRI-NIFA Fellowship Grant

K.E. Papa Outstanding Ph.D. Student, Department of Plant Pathology, UGA
 Student Travel Award, Graduate School, UGA
 Second Place Oral Presentation, Broadus Browne Competition, UGA
2011: 11th I.E. Melhus Graduate Student Symposium Award, APS
 Dissertation Completion Award, Graduate School, UGA
 Grants-in-Aid-of-Research Award, Sigma Xi
 R.J. Tarleton Fellowship, APS
 Student Travel Award, Graduate School, UGA
2010: Outstanding Graduate Teaching Assistant, Department of Plant Pathology, UGA
2009: C. Lee Campbell Student Travel Award, APS
 First Place Student Presentation, Georgia Association of Plant Pathologists
2008: First Place Graduate Thesis Award, UCM
2007: Microbiology Research Award, Association of Southeastern Biologists
 Outstanding Graduate Student, Department of Biology, UCM
 Quarterman-Keever Poster Award, Southeastern Ecological Society of America
2006: Willard North Graduate Award for Research, UCM
2005: Dan Cooper Memorial Scholarship, Iowa State Horticulture Society
2004: Midwest Aquatic Plant Management Society
2000: “Iowa’s Promise...Our Youth” Grant

Invited Seminars and Symposia (11 total):

2017: 16th International Sclerotinia Workshop, Uberlandia, Brazil (*two invited talks*)
 Department of Plant Pathology, Ohio State University, Wooster, OH (*and workshop*)
 Department of Microbiology and Plant Pathology, Iowa State University, Ames, IA
2016: Schroth Faces of the Future Symposium, Epidemiology, APS Meeting, Tampa, FL
 Department of Plant Pathology, UNL, Seminar Series, Lincoln, NE
2015: Department of Plant and Environmental Sciences, Clemson University, Clemson, SC
 Department of Biology and Agriculture, University of Central Missouri, Warrensburg, MO
2014: Plant Science Retreat, University of Nebraska, Nebraska City, NE
 Special Session on Genotyping-by-Sequencing, APS Meeting, Austin, TX
2013: Institute of Agriculture and Natural Resources, University of Nebraska, Lincoln, NE
2011: 11th I.E. Melhus Graduate Student Symposium, Epidemiology, APS Meeting, Honolulu, HI

Teaching Experience (14 years experience):

- Success in the Sciences, UNL, Summer 2018
- Ecology and Management of Plant Pathogens, UNL, Spring 2016, 2017, 2018, and every spring
- Plant Diseases Across Nebraska, UNL, Summer 2016, 2017
- Population Genetics and Advanced Epidemiology in R, UNL, Summer 2016
- Disease Dynamics and Evolution, UNL, Spring 2016
- Mycology, Lab TA and guest lecturer, UGA, 2009 and 2010
- Introductory Plant Pathology, Lab TA, UGA, Fall 2008
- Botany, Lab TA, UCM, Fall 2006
- Anatomy and Physiology, Lab TA, UCM, Fall 2006
- Math Learning Center, Tutor, UCM, Spring 2005
- Introductory Algebra, Instructor for 2 sections, UCM, Fall 2004

Workshops in R (7 total):

- **Everhart, S.E.**, N.G. Gambhir, and Z.N. Kamvar. 2018. Intro to R for Plant Pathologists
Most recent workshop website: everhartlab.github.io/IntroR_Workshop
 - 64 attendees, International Congress of Plant Pathology, Boston, MA, August 28, 2018
 - 45 attendees, University of Nebraska, Lincoln, NE, June 27, 2018;
 - 20 attendees, Ohio State University, Wooster, OH, October 16, 2017;
 - 22 attendees, APS North Central Division Meeting in Champaign, IL, June 14, 2017;
 - 56 attendees, University of Nebraska, Lincoln, NE, May 24, 2017;
- Grünwald, N.J., Z.N. Kamvar, and **S.E. Everhart**. 2014. Population Genetics in R.
grunwaldlab.github.io/Population_Genetics_in_R
 - 40 attendees, Oregon State University, May 2014.
 - 60 attendees, APS National Meeting in Austin, TX, August 2014

Students and Postdoctoral Scholars in my lab at UNL (18 total):

- Postdoctoral Scholars – Margarita Marroquin-Guzman, 2017-**present**; Zhian N. Kamvar, 2017-2018; Thomas J.J. Miorini, 2016-2018; B. Sajeewa Amaradasa, 2014-2016
- Graduate Students – Karen Ferreira Da Silva, Ph.D., 2018-**present**; Edgar Lopez, Ph.D., 2016-**present**; Nikita Gambhir, Ph.D., 2015-**present**
- Undergraduate Students – Rachel Persson 2018-**present**; Isabel Chavez, 2017-2018; Audrey Vega, 2017-2018; Alex Johnson, 2017-2018; Anthony Pannullo, 2015-2017; Morgan Thompsen, 2016; Josh Hanson, 2014-2017; Sarah Campbell, 2014-2016
- Graduate Interns and Rotations – Callie Braley, D.PH., Summer Internship 2018-**present**; Bridget Tripp, Ph.D. Complex Biosystems, Fall 2015
- Undergraduate Intern – Flavio Nunes da Silva, Brazilian CNPq Fellowship Program, 2015

Professional Service (since 2014):

- Professional positions – Section Editor for *Tropical Plant Pathology* (2018-**present**), APS Annual Meeting Advisory Board (2016-**present**), Vice-Chair of the Epidemiology Committee of APS (2018-**present**) Associate Editor for *Ciencias Rural* (2014-2015), Vice-Chair, Chair, and Immediate Past Chair of the Mycology Committee of APS (2013-2016)
- Departmental/IANR Committees at UNL – Curriculum Committee (2018**present**); Co-Chair, *Ad hoc* Plant Pathology Graduate Program Proposal Committee (2018**present**); CASNR Web Framework (2015**present**); Chair, Website Committee (2015-2018); Academic Review Committee for Department of Entomology at UNL; (2016); Hired and supervised departmental staff: Jimin Kamvar, *Digital Communications Liaison*, October 2017–December 2017.
- Ad hoc peer reviews for – *Annals of Botany*, *Crop Protection*, *Ciencias Rural*, *European Journal of Plant Pathology*, *Journal of Phytopathology*, *PeerJ*, *Plant Disease*, *PLOS ONE*, *Phytopathology*, *Scientia Agricola*, *Tropical Plant Pathology*
- Grant panels – USDA Foundational Program for Pests and Beneficial Species (December 2017), USDA External Review of Research Plans (January 2017)

Peer-Reviewed Publications (27 total):

*Co-first author; **Corresponding author, †Undergraduate

1. Kamvar, Z., and **S.E. Everhart**** . 2018. Something in the agar does not compute: On the discriminatory power of mycelial compatibility in *Sclerotinia sclerotiorum*. *Tropical Plant Pathology*. *In press*. Preprint available at *PeerJ Preprints* 6:e26670v1
2. Pannullo†, A., Kamvar, Z., Miorini, T.J.J., and **S.E. Everhart**** . 2018. Phenotypic and genotypic variation of *Sclerotinia sclerotiorum* from North and South America. *Tropical Plant Pathology*. *Accepted*. Preprint available at *PeerJ Preprints* 6:e26600v1
3. Bogo, A., C.C. Comparin, R.M.V. Sanhueza, P. Ritschel, R.T. Casa, F.N. Silva, and **S.E. Everhart**. 2018. Characterization of two *Neofabraea* species, *Neofabraea actinidiae* and *N. brasiliensis*, that are causal agents of apple bull's-eye rot in southern Brazil. *Canadian Journal of Plant Pathology*. *Pre-print*: <https://doi.org/10.1080/07060661.2017.1421588>
4. Kamvar, Z., Amaradasa, B.S., R. Jhala, S. McCoy, J.R. Steadman, and **S.E. Everhart**** . 2017. Population structure and phenotypic variation of *Sclerotinia sclerotiorum* from dry bean in the United States. *PeerJ*. 5:e4152 <https://doi.org/10.7717/peerj.4152> **“Top 5 Most Viewed Articles of 2017”** in *PeerJ*'s sections on Agriculture Science, Genetics, and Mycology
5. Dowling, M., G. Schnabel, H. Boatwright†, and **S.E. Everhart**** . 2017. Novel gene-sequence markers for isolate tracking within *Monilinia fructicola* lesions. *Pest Management Science* 73:1822-1829. DOI:10.1002/ps.4544.
6. Grünwald, N.J., **S.E. Everhart**, B.J. Knaus, and Z.N. Kamvar. 2017. Best practices for population genetic analyses. *Phytopathology*. 107:1000–1010. **Sept. 2017 Phytopathology Cover Image**
7. Miorini, T.J.J., C.G. Raetano, and **S.E. Everhart**** . 2017. Control of white mold of dry bean and residual activity of fungicides applied by chemigation. *Crop Protection*. 94:192-202.
8. Amaradasa, B.S., and **S.E. Everhart**** . 2016. Effects of sublethal fungicides on mutation rates and genomic variation in fungal plant pathogen, *Sclerotinia sclerotiorum*. *PLoS ONE*. 11(12): e0168079. DOI 10.1371/journal.pone.0168079.
9. de Bem, B.P., A. Bogo, **S.E. Everhart**, R.T. Casa, M.J. Goncalves, J.L. Marcon, L.R. Rufato, F.N. Silva, R. Allebrandt, I.C. da Cunha. 2016. Effect of four training systems on the temporal dynamics of downy mildew in two grapevine cultivars in southern Brazil. *Tropical Plant Pathology*. DOI 10.1007/s40858-016-0110-8.
10. Dowling, M., P.K. Bryson, H. Boatwright†, J.R. Wilson, Z. Fan, G. Schnabel, **S.E. Everhart**, and P. Brannen. 2016. Effect of fungicide application on *Monilinia fructicola* population diversity and transposon movement. *Phytopathology* 106:1504–1512.
11. Dugan, F.M., and **S.E. Everhart**. 2016. Cryptic species: A leitmotif of contemporary mycology has challenges and benefits for plant pathologists. *Plant Health Progress* 17:250-253. DOI10.1094/PHP-RV-16-0046. **Feb. 2017 Plant Health Progress Editor's Pick**
12. Tabima J.F., **S.E. Everhart**, M.M. Larsen, A.J. Weisberg, Z.N. Kamvar, M.A. Tancos, C.D. Smart, J.H. Chang, and N.J. Grünwald. 2016. Microbe-ID: An open source toolbox for microbial genotyping and species identification. *PeerJ* 4:e2279 DOI 10.7717/peerj.2279.
13. Chen, F., **S.E. Everhart***, P.K. Bryson, C.L., X. Song, X.L., G. Schnabel. 2015. Fungicide induced transposon movement in *Monilinia fructicola*. *Fungal Genetics and Biology* 85:38–44.
14. de Bem, B.P., Bogo, A., **S.E. Everhart**, R.T. Casa, M.J. Goncalves, J.L.M. Filho, and I.C. da Cunha. 2015. Effect of Y-trellis and vertical shoot positioning training systems on downy mildew

- and botrytis bunch rot of grape in highlands of southern Brazil. *Scientia Horticulturae* 185:162–166.
15. **Everhart, S.E.**, and H. Scherm. 2015. Clonal disease foci of *Monilinia fructicola* during brown rot epidemics within peach tree canopies. *Phytopathology*. 105:542–549.
 16. Schnabel, G., F. Chen, **S.E. Everhart**, W.C. Bridges and X.L. Liu. 2014. Studies on sensitivity reduction in solo and mixture treatments and fungicide-induced mutagenesis in *Monilinia fructicola*. In: H.W. Dehne, H.B. Deising, U. Gisi, B. Fraaije, U. Gisi, D. Hermann, A. Mehl, E.C. Oerke, P.E. Russel, G. Stammler, K.H. Kuck, H. Lyr (Eds). “Modern Fungicides and Antifungal Compounds”, Vol. VII, pp 263-268. 2014 Deutsche Phytomedizinische Gesellschaft, Braunschweig, ISBN: 978-3-941261-13-6.
 17. **Everhart, S.E.**, A. Askew, L. Seymour, and H. Scherm. 2013. Spatio-temporal patterns of pre-harvest brown rot epidemics within individual peach tree canopies. *European Journal of Plant Pathology* 135:499–509.
 18. **Everhart, S.E.**, A. Askew, L. Seymour, T.C. Glenn, and H. Scherm. 2012. Spatial patterns of brown rot epidemics and development of microsatellite markers for analyzing fine-scale genetic structure of *Monilinia fructicola* populations within peach tree canopies. Online. *Plant Health Progress* doi:10.1094/PHP-2012-0723-04-RS.
 19. **Everhart, S.E.**, A. Askew, L. Seymour, I.J. Holb, and H. Scherm. 2011. Characterization of three-dimensional spatial aggregation and association patterns of brown rot symptoms within intensively mapped sour cherry trees. *Annals of Botany* 108:1195–1202.
 20. **Everhart, S.E.**** 2010. Collection and identification of grapevines (*Vitis*) from the tree canopy of select forests in the southeastern United States. *Castanea* 75: 141–149.
 21. Keller, H.W., and **S.E. Everhart** 2010. Importance of Myxomycetes in biological research and teaching. *Fungi* 3(1):13–27.
 22. **Everhart, S.E.**, J.S. Ely, and H.W. Keller. 2009. Evaluation of tree canopy epiphytes and bark characteristics associated with corticolous myxomycetes. *Botany* 87:509–517.
 23. Keller, H.W., **S.E. Everhart**, M. Skrabal, and C.M. Kilgore. 2009. Tree canopy biodiversity in temperate forests: Exploring islands in the sky. *Southeastern Biology* 56:52–74.
 24. **Everhart, S.E.**, and H.W. Keller. 2008. Influence of bark pH on the occurrence and distribution of tree canopy myxomycete species. *Mycologia* 100:191–204.
 25. **Everhart, S.E.**, and H.W. Keller. 2008. Life history strategies of corticolous myxomycetes: The life cycle, fruiting bodies, plasmodial types, and taxonomic orders. *Fungal Diversity* 29:1–16.
 26. Keller, H.W., and **S.E. Everhart** 2008. Myxomycete species concepts, monotypic genera, the fossil record, and additional examples for good taxonomic practice. *Revista Mexicana de Micologia* 27:9–19.
 27. Keller, H.W., C.M. Kilgore, **S.E. Everhart**, G. Carmack, C. Crabtree, and A. Scarborough. 2008. Myxomycete plasmodia and fruiting bodies: Unusual occurrences and user friendly study techniques. *Fungi* 1:24–37.
 28. Kilgore, C.M., H.W. Keller, **S.E. Everhart**, A. Scarborough, K. Snell, M. Skrabal, C. Pottorff, and J.S. Ely. 2008. Tree canopy research and student experiences using the double rope climbing method. *Journal of Botanical Research Institute of Texas* 2:1309–1336.

Publications in review and in preparation:

29. Miorini, T.J.J., R. Higgins, C.G. Raetano, J.R. Steadman, and **S.E. Everhart****. 201X. Variation in pathogen aggression and cultivar performance against *Sclerotinia sclerotiorum*. *Tropical Plant Pathology*. Under revision. Preprint available at *PeerJ Preprints* 6:e26622v1
30. Miorini, T.J.J., E.N. Lopez, N.K. Gambhir, A. Pannullo[†], and **S.E. Everhart****. 201X. Comparison of methods used to assess fungicide sensitivity in *Sclerotinia sclerotiorum*. *Fungal Genetics and Biology*. In preparation.
31. Miorini, T.J.J., R. Higgins, J.R. Steadman, and **S.E. Everhart****. 201X. Fungicide sensitivity of *Sclerotinia sclerotiorum* isolates from Brazil, Argentina, and the U.S.A. *Plant Disease*. In preparation.
32. Ajayi, O.O., **S.E. Everhart**, P.J. Brown, A.U. Tenuta, A.E. Dorrance, and C. Bradley. 201X. Genetic structure of *Rhizoctonia solani* AG-2-2IIIB from soybean in Illinois, Ohio, and Ontario. *Phytopathology*. In preparation.
33. Dale, A.L., N. Feau, **S.E. Everhart**, G. Bilodeau, B. Dhillon, J. Tabima, C. Brasier, N. Grünwald, R.C. Hamelin. 201X. Mitotic recombination and a two-speed genome drive evolution in asexual lineages of the sudden oak death pathogen *Phytophthora ramorum*. In review.
34. Matczyszyn, J., **S.E. Everhart**, K. Powers, T. Harris, and T.O. Powers. 201X. Diversity and Distribution of Criconematidae nematodes in Eastern North America. In preparation.

Books, Chapters, Proceedings, R Packages, and Magazine Articles:

1. Gambhir, N., **S. Everhart**, S. Kodati, & A. Adesemoye. 2018. Fungicide Resistance: Risk and Management. *SoybeanNebraska Magazine*, Spring 2018, Page 22
2. Kodati, S., A. Adesemoye, N. Gambhir, & **S. Everhart**. 2018. Rhizoctonia Diseases in Soybean. *SoybeanNebraska Magazine*, Spring 2018, Page 23
3. Keller, H.W., **S.E. Everhart**, and C.M. Kilgore. 2017. The Myxomycetes: Biology, life cycle, genetics and reproduction. In: Stephenson, S. and C. Lado (eds) “Myxomycetes: Biology, Systematics, Biogeography and Ecology”, Elsevier, Atlanta, GA.
4. Miorini, T.J., A. Pannullo[†], T. Hornby[†], R. Higgins, **S.E. Everhart****, and J.R. Steadman. 2017. Phenotypic and genotypic characterization of relevant *Sclerotinia sclerotiorum* isolates. *Bean Improvement Cooperative*.
5. **Everhart, S.E.****, B.S. Amaradasa, R. Jhala, R. Higgins, and J.R. Steadman. 2016. Population structure and fungicide sensitivity of 366 *Sclerotinia sclerotiorum* isolates from dry common bean. *Bean Improvement Cooperative* 59:131–132.
6. Kamvar, Z.N., Tabima, J.F., **Everhart, S.E.**, Brooks[†], J.C., Krueger-Hadfield, S.A., Sotka, E. and Grünwald, N.J., 2016. Package ‘poppr’. <https://cran.r-project.org/web/packages/poppr>
7. Grünwald, N.J., Z.N. Kamvar, and **S.E. Everhart**. 2015. Population Genetics in R. Online book: http://grunwaldlab.github.io/Population_Genetics_in_R
8. **Everhart, S.E.**, T.F. Tabima, and N.J. Grünwald. 2014. *Phytophthora ramorum*. In: Dean, R.A., A. Lichens-Park, and C. Kole (eds) “Genomics of Plant Associated Fungi and Oomycetes”, Springer, New York, NY. Pp. 159–174.
9. **Everhart, S.E.**. 2008. Edible, avoidable, and artistic fungi for summer and fall. *The Iowa Horticulturist* 24(2): 22–23.
10. **Everhart, S.E.**. 2008. Edible and avoidable fungi for spring. *The Iowa Horticulturist* 24(1): 22–23.
11. **Everhart, S.E.**. 2007. Smooth patch on oak trees. *The Iowa Horticulturist* 23(3): 17.

12. **Everhart, S.E.**. 2006. Slime invaders on your lawn. *The Iowa Horticulturist* 22(2): 18–20.
13. Keller, H.W. and **S.E. Everhart**. 2006. Myxomycetes (true slime molds): Educational sources for students and teachers (Part I and II). *Inoculum* 57(3): 1–2; 57(4): 4–5.
14. **Everhart, S.E.** 2002. Daylily rust in Iowa. *The Iowa Horticulturist* 19(1): 18–20.
15. **Everhart, S.E.**. 2000. Wildflower demonstration garden. *The Iowa Horticulturist* 17(1): 14–15.
16. **Everhart, S.E.**. 1999. White pines in Iowa. *The Iowa Horticulturist* 15(2): 10.

Poster and Oral Presentations (28 presented by member of Everhart Lab):

◊ = presenting author and member of Everhart Lab

1. Gambhir[◊], N., S. Kodati, A.O. Adesemoye, and **S.E. Everhart**. 2018. Fungicide sensitivity of *Rhizoctonia* spp. isolated from soybean fields in Nebraska. ICPP / APS National Meeting
2. Marroquin-Guzman[◊], M.R., C. Proctor, J. McMechan, R. Werle, A.O. Adesemoye, and **S.E. Everhart**. 2018. Soil fungal diversity during a soybean-cover crop rotation using community sequencing. ICPP / APS National Meeting
3. Kamvar, Z.N. and **S.E. Everhart**[◊]. 2018. The open road: A case study of reproducible research in plant pathology. ICPP / APS National Meeting
4. Kamvar, Z.N., B.S. Amaradasa, R. Jhala, S.B. McCoy, J.R. Steadman, and **S.E. Everhart**[◊]. 2018. White mold/dry bean: Population structure and phenotypic variation of *Sclerotinia sclerotiorum* from dry bean in the USA. ICPP / APS National Meeting
5. Gazis, R.O., **S.E. Everhart**, A. Graves, Z.N. Kamvar, S.N. Trigiano, S.J. Seybold, and D. Hadziabdic. 2018. When natives become invasive: Population genetic signatures following range expansion in members of thousand cankers disease complex. ICPP / APS National Meeting
6. Arneson, N., L.J. Giesler, R. Werle, and **S.E. Everhart**. 2018. Effect of soil-applied protoporphyrinogen oxidase inhibitor herbicides on root rot severity of soilborne pathogens in soybean [*Glycine max* (L.) merr.]. ICPP / APS National Meeting
7. Gambhir[◊], N., Z.N. Kamvar, and **S.E. Everhart**. 2018. Genomic signatures of sub-lethal fungicide stress in *Sclerotinia sclerotiorum*. ICPP / APS National Meeting
8. Nieto-Lopez[◊], E., T.J.J. Miorini, and **S.E. Everhart**. 2018. Fungicide sensitivity of 42 *Sclerotinia sclerotiorum* isolates in the North Central U.S. and determination of discriminatory concentrations. APS North Central Division Meeting
9. Stengel, A., J. Herr, E. Jeske, V. Jin, M. Schmer, **S.E. Everhart**, R. Drijber. 2018. Insights from 40 years of maize cropping: Crop diversity shapes soil nutrient pools and drives bacterial community structure. Soil Science Society of America National Meeting
10. **Everhart**[◊], **S.E.**, & A.O. Adesemoye. 2018. Fungicide resistance in *Rhizoctonia solani* and implications for soybean fields in Nebraska. *Research Update*. Nebraska Soybean Board, Columbus, NE, January 10, 2018.
11. Arneson, N., L.J. Giesler, R. Werle, and **S.E. Everhart**. 2017. Effect of soil-applied sulfentrazone and flumioxazin on soybean seedling disease severity under field conditions. North Central Weed Science Society Annual Meeting.
12. **Everhart**[◊], **S.E.**, Z.N. Kamvar, B.S. Amaradasa, T.J.J. Miorini, R. Jhala, A. Pannullo, R. Higgins, J.R. Steadman. 2017. *Sclerotinia sclerotiorum* in North America: Recent disease outbreaks and variability of populations across the United States and Mexico. 16th International Sclerotinia Workshop in Uberlandia, Brazil. **Invited by Dr. David Jaccoud Filho.**
13. **Everhart**[◊], **S.E.**, N.K. Gambhir, Z.N. Kamvar. 2017. Effect of sublethal fungicide exposure on genomic variation in *Sclerotinia sclerotiorum*. 16th International Sclerotinia Workshop in Uberlandia, Brazil. **Invited by Dr. Eduardo Mizubuti.**

14. **Everhart[◇], S.E.**, 2017. Causes and consequences of population genetic variation in *Sclerotinia sclerotiorum*. Seminar in Department of Plant Pathology and Microbiology at Iowa State University. **Invited by Dr. Mark Gleason.**
15. **Everhart[◇], S.E.**, 2017. Causes and consequences of population genetic variation in *Sclerotinia sclerotiorum*. Seminar in Department of Plant Pathology at Ohio State University, Wooster, OH. **Invited by Dr. Anne Dorrance.**
16. Gambhir[◇], N., Z.N. Kamvar, and **S.E. Everhart**. 2017. Effects of sublethal fungicide stress on genomes of *Sclerotinia sclerotiorum*. APS National Meeting
17. Gambhir[◇], N., Z.N. Kamvar, and **S.E. Everhart**. 2017. Genomic alterations in *Sclerotinia sclerotiorum* after sublethal exposure to a mitosis-inhibiting fungicide. APS North Central Division Meeting
18. Kamvar[◇], Z.N., **S.E. Everhart**, and N. Grünwald. 2017. I think we're a clone now: Factors influencing inference of clonality in diploid populations. APS National Meeting
19. Kodati, S., N. Gambhir[◇], **S.E. Everhart**, and A.O. Adesemoye. 2017. Prevalence and pathogenicity of *Rhizoctonia* spp. from soybean in Nebraska. APS National Meeting.
20. Miorini[◇], T.J., A. Pannullo[†], J.R. Steadman, and **S.E. Everhart**. 2017 Fungicide sensitivity and population structure of *Sclerotinia sclerotiorum* isolates from Argentina, Brazil, and USA. APS National Meeting
21. Miorini[◇], T.J.J., **S.E. Everhart**, and J. Steadman. 2017. Fungicide sensitivity of *Sclerotinia sclerotiorum* isolates from Brazil, Argentina, and the USA. APS National Meeting
22. Nieto-Lopez[◇], E.H., and **S.E. Everhart**. 2017. Fungicide sensitivity of *Sclerotinia sclerotiorum* from soybean in the North Central United States. APS North Central Division Meeting
23. Pannullo[◇], A., T.J.J. Miorini, Z. Kamvar, and **S.E. Everhart**. 2017. Population genetic diversity of *Sclerotinia sclerotiorum* populations from Brazilian soybean. APS North Central Division
24. Stengel, A., S. Ramirez II, E.S. Jeske, V.L. Jin, J. Cui, **S.E. Everhart**, J. Herr, and R Drijber. 2017. Nitrogen and crop rotation as drivers of the maize-associated soil microbiome. *Argonne Soil Metagenomics Meeting*, Chicago, IL
25. Amaradasa[◇], B.S., and **S.E. Everhart**. 2016. Sub-lethal fungicides induce microsatellite mutation in *Sclerotinia sclerotiorum*. *Phytopathology* 106:S4.139.
26. Amaradasa[◇], B.S., and **S.E. Everhart**. 2016. Sub-lethal fungicides induce microsatellite and AFLP marker mutation in *Sclerotinia sclerotiorum*. *Phytopathology* 106:S4.184.
27. Dowling, M., H. Boatwright[†], G. Schnabel, P. Bryson, J. Wilson, Z. Fan, **S.E. Everhart**, and P. Brannen. 2016. Effect of fungicide applications on *Monilinia fructicola* population diversity and transposon movement. *Phytopathology* 106:S4.62.
28. **Everhart[◇], S.E.**, and B.S. Amaradasa. 2016. Fungicide stress induces genome mutation in *Sclerotinia sclerotiorum*. *Phytopathology* 106:S4.169.
29. **Everhart[◇], S.E.**, R. Higgins, and J.R. Steadman. 2016. “Sources of white mold resistance derived from wide crosses in common bean and progress in characterization of relevant pathogen isolates. National Sclerotinia Initiative Meeting.
30. Gambhir[◇], N., A. Pannullo[†], S. Campbell[†], B.S. Amaradasa, R. Jhala, J. Steadman, and **S.E. Everhart**. 2016. Comparison of four methods for fungicide sensitivity determination of *Sclerotinia sclerotiorum*. *Phytopathology* 106:S4.188.
31. T.J. Miorini[◇], C.G. Raetano, and **S.E. Everhart**. 2016. Residual effect of fungicides applied by chemigation for white mold control in dry bean. *Phytopathology* 106:S4.190.
32. T.J. Miorini[◇], R. Werle, A. Stavievski, C.G. Raetano, and **S.E. Everhart**. 2016. Evaluation of residual fungicide in soybean leaves using analytical chemical quantification and *Sclerotinia sclerotiorum* bioassay. *Phytopathology* 106:S4.189.

33. Amaradasa[◇], B.S., and **S.E. Everhart**. 2015. Sub-lethal doses of fungicide induce resistance emergence in *Sclerotinia sclerotiorum*. *Phytopathology*. 105:S4.7.
34. **Everhart[◇], S.E.**, R. Jhala, B.S. Amaradasa, R. Higgins, J.R. Steadman. 2015. Worldwide population structure of *Sclerotinia sclerotiorum* from cultivated common bean. *Phytopathology*. 105:S4.41.
35. **Everhart, S.E.**, B.J. Knaus, A. Kanaskie, W. Sutton, P. Reeser, A.L. Dale, R.C. Hamelin, E. Hansen, and N.J. Grünwald. 2014. Exploring the use of genotyping-by-sequencing to characterize the forest epidemic of *Phytophthora ramorum* in Oregon. *Phytopathology* 104:S3.153. **Invited seminar for the APS Special Session on Genotyping-by-Sequencing.**
36. **Everhart, S.E.**, M.M. Larsen, A. Kanaskie, and N.J. Grünwald. 2014. Early detection of *P. ramorum* lineages in Oregon forests using genetic markers. *USDA Forest Health Management Working Group Meeting*, Jacksonville, FL.
37. Dale, A.L., **S.E. Everhart**, N. Feau, G.L. Bilodeau, N.J. Grünwald, and R.C. Hamelin. 2013. Genome-wide patterns of diversity in four lineages of the sudden oak death pathogen, *Phytophthora ramorum*. *Phytopathology*. 103:S2.32.
38. **Everhart, S.E.** 2013. *Phytophthora ramorum* blight: where is it now? *Conference on Soil Borne Plant Pathogens*, Corvallis, OR.
39. **Everhart, S.E.** and Scherm, H. 2013. Clonal population foci of *Monilinia fructicola* during epidemics within peach tree canopies. (Abstr.). *Acta Phytopathologica Sinica* 43:173.
40. **Everhart, S.E.**, M.M. Larsen, N.J. Grünwald. 2013. Where is *Phytophthora ramorum* now? An update on clonal populations in the U.S. *Phytopathology*. 103:S2.41
41. Scherm, H., and **Everhart, S.E.** 2013. Spatial, temporal, and population aspects of epidemics in fruit tree canopies. Pages 14-15 in: *Proceedings of the 4th Brazilian Workshop of Plant Disease Epidemiology*, Fed. Univ. Parana, Curitiba, Brazil.
42. Tabima, J.F., **S.E. Everhart**, M.M. Larsen, Z.N. Kamvar, and N.J. Grünwald. 2013. Phytophthora-ID 2.0: Novel open source tools for genotype and species identification of *Phytophthora* spp. *Center for Genome Research and Biocomputing Conference*, Corvallis, OR.
43. **Everhart, S.E.**, M.M. Larsen, A. Kanaskie, and N.J. Grünwald. 2014. Early detection of *P. ramorum* lineages in Oregon forests using genetic markers. *USDA Forest Health Management Working Group Meeting*, Jacksonville, FL.
44. Scherm, H., and **Everhart, S.E.** 2013. Spatial, temporal, and population aspects of epidemics in fruit tree canopies. (Abstr.) Pages 14-15 in: *Proceedings of the 4th Brazilian Workshop of Plant Disease Epidemiology*, Fed. Univ. Parana, Curitiba, Brazil.
45. **Everhart, S.E.** and Scherm, H. 2013. Clonal population foci of *Monilinia fructicola* during epidemics within peach tree canopies. (Abstr.). *Acta Phytopathologica Sinica* 43(Suppl.):173.
46. Dale, A.L., **S.E. Everhart**, N. Feau, G.L. Bilodeau, N.J. Grünwald, and R.C. Hamelin. 2013. Genome-wide patterns of diversity in four lineages of the sudden oak death pathogen, *Phytophthora ramorum*. *Phytopathology*. 103(Suppl. 2):S2.32.
47. **Everhart, S.E.** 2013. *Phytophthora ramorum* blight: where is it now? *Conference on Soil Borne Plant Pathogens*, Corvallis, OR.
48. **Everhart, S.E.**, M.M. Larsen, and N.J. Grünwald. 2013. Where is *Phytophthora ramorum* now? An update on clonal populations in the U.S. *Phytopathology*. 103(Suppl. 2):S2.41.
49. Tabima, J.F., **S.E. Everhart**, M.M. Larsen, Z.N. Kamvar, and N.J. Grünwald. 2013. Phytophthora-ID 2.0: Novel open source tools for genotype and species identification of *Phytophthora* spp. *Center for Genome Research and Biocomputing Conference*, Corvallis, OR.
50. **Everhart, S. E.** and H. Scherm. 2012. Fine-scale genetic structure of *Monilinia fructicola* populations within peach tree canopies. *Phytopathology* 102:S436.

51. **Everhart, S.E.**, A. Askew, L. Seymour, I.J. Holb, and H.Scherm. 2011. Spatial distribution of brown rot symptoms and fine-scale genetic structure of populations of *Monilinia* sp. within and among stone fruit tree canopies. *Phytopathology* 101:S221.
52. **Everhart, S.E.**, H. Scherm, A. Askew, L. Seymour, and I.J. Holb. 2010. Characterization of 3-D spatial association and aggregation patterns of brown rot symptoms within intensively mapped fruit trees. *Proceedings of the 6th International Workshop on Functional-Structural Plant Models*.
53. —. 2009. Three-dimensional spatial patterns of brown rot symptoms within sour cherry tree canopies in Hungary. *Phytopathology* 99:S33.
54. **Everhart, S.E.** 2009. 3D Spatial patterns of brown rot symptoms within sour cherry tree canopies in Hungary. *Georgia Association of Plant Pathologists*.
55. Scherm, H., **S.E. Everhart**, A. Askew, L. Seymour, and I.J. Holb. 2009. Spatial patterns of brown rot symptoms in individual, intensively mapped cherry trees. Pages 143-145 in: *Proc. 10th Intl. Epidemiol. Workshop*. Gadoury, Seem, Moyer, and Fry, eds. NY State Ag. Experiment Station, Geneva, NY. ISBN 0-9676507-7-1.
56. **Everhart, S.E.** 2009. Tree canopy exploration: The last frontier of biological research on earth. *International Master Gardener's Conference*, Las Vegas, NV. Keynote speaker.
57. **Everhart, S.E.** 2007. Species Assemblages and Distribution of Corticolous Myxomycetes in the Tree Canopy of Selected Forests. *Department of Plant Pathology*, UGA.
58. **Everhart, S.E.** 2008. Influence of bark pH on the assemblage and distribution of corticolous myxomycetes in the tree canopy in Great Smoky Mountains National Park. *Mid-Atlantic States Mycological Conference*.
59. **Everhart, S.E.** 2008. Vertical variation in bark characteristics and epiphyte cover on distribution patterns of corticolous myxomycetes (true slime molds) in the tree canopy. *Mid-Atlantic States Mycological Conference*.
60. **Everhart, S.E.** 2007. Tree Canopy Myxomycetes (True Slime Molds): Distribution patterns and species assemblages on trees and grapevines in temperate forests. *Sigma Xi Annual Meeting and Student Research*.
61. **Everhart, S.E.** 2007. The hunt for “biological jewels of nature” in the Great Smoky Mountains National Park. *Iowa State University Shade Tree Short Course*.
62. **Everhart, S.E.**, H.W. Keller, and J.S. Ely. 2007. Corticolous myxomycetes (true slime molds): species assemblages and distribution in the tree canopy of selected forests of Kentucky and Tennessee. *Mycological Society of America Annual Meeting and Foray*
63. **Everhart, S.E.**, H.W. Keller, and J.S. Ely. 2007. Quantitative analysis of bark characteristics and epiphyte cover on distribution patterns of corticolous myxomycetes (true slime molds) in the tree canopy. *Mycological Society of America*.
64. **Everhart, S.E.**, H.W. Keller, and J.S. Ely. 2007. Ecology of canopy myxomycetes (true slime molds) on trees and grapevines (*Vitis aestivalis* and *V. vulpina*). *Southeastern Biology* 54 (2).
65. **Everhart, S.E.**, H.W. Keller, and J.S. Ely. 2007. Role of bark characteristics and epiphyte cover in the abundance, distribution, and succession of corticolous myxomycetes (true slime molds). *Southeastern Biology* 54 (2).
66. Keller, H.W., **S.E. Everhart**, C.M. Kilgore, A. Scarborough, and G. Carmack. Myxomycete plasmodial tracks and fruiting bodies on animal skulls, fungi, myxomycetes, rocks, spiders, and canopy bark of living trees. *Mycological Society of America*.

67. Keller, H.W., **S.E. Everhart**, C.M. Kilgore, A. Scarborough, and G. Carmack. 2007. Myxomycetes, the true slime molds of Kentucky: new species and records, with field and laboratory observations of plasmodial tracks on a canine skull. *Southeastern Biology* 54 (2).
68. **Everhart, S.E.**, and H.W. Keller. 2006. Ecology of canopy myxomycetes on grapevines. *Transactions of the Missouri Acad. of Science*, Collegiate Division, Vol. 42.
69. **Everhart, S.E.**, H.W. Keller, J.S. Ely, and S. Wilson. 2006. Slime, skulls, and the giant grapevine: ecology of canopy myxomycetes (true slime molds) on trees and grapevines (*Vitis aestivalis* and *V. vulpina*). *Sigma Xi Annual Meeting and Student Research Conference*, Program of Events (Program Update Addendum). EB-14. Page 12.
70. Keller, H.W., and **S.E. Everhart**. 2006. Ecology of canopy myxomycetes on grapevines. *Transactions of the Missouri Acad. of Science*, Collegiate Division, Vol. 42.
71. Keller, H.W., and **S.E. Everhart**. 2006. Myxomycetes (true slime molds): educational sources for students and teachers. *Transactions of the Missouri Acad. of Science*, Collegiate Division, Vol. 42.
72. **Everhart, S.E.** 2005. Spatial distribution of gopher mounds of *Geomys bursarius* in Cayler Prairie State Preserve. *Lakeside Lab Public Address*.
73. **Everhart, S.E.** 2001. Affect of Benomyl fungicide on *Sphaeropsis sapinea*, the causal agent of Sphaeropsis shoot blight and canker. *Undergraduate Research Scholars Symposium*.

TEACHING – past 5 years

Courses Taught

2-year average CASNR Calculated FTE: 25.11%

2018 CASNR Calculated FTE: 24.18% Instructional and 8.27% Advising

- **Ecology and Management of Plant Pathogens (PLPT 802)** lead instructor (60% credit), Spring 2018, 3 credit hours, 15 graduate students
- **Success in the Sciences (PLPT 896 *Special Topics*)** lead instructor (100% credit), Summer 2018, 2 credit hours, 7 graduate students (1-audit)
- **Hands-On Instructional Design (PLPT 896 *Special Topics*)** lead instructor (100% credit), Summer 2018, 2 credit hours, 2 graduate students enrolled

2017 CASNR Calculated FTE: 11.53% Instructional and 6.24% Advising

- **Ecology and Management of Plant Pathogens (PLPT 802)** lead instructor (40% credit), spring 2017, 3 credit hours, 17 graduate students
- **Field Disease Tour (PLPT 891)** co-instructor (50% credit), fall 2017, 1 credit hour, 14 graduate students

2016 CASNR Calculated FTE: 30.06% Instructional and 5.29% Advising

- **Advanced Epidemiology and Population Genetics in R (PLPT 892)** taught in summer 2016 (100% credit), 1 credit hour, 2 graduate students (28 others in attendance)
- **Disease Dynamics and Evolution (PLPT 892/496)** taught spring 2016 (100% credit), 3 credit hours, 2 graduate students and 4 undergraduates
- **Ecology and Management of Plant Pathogens (PLPT 802)** co-taught (30% credit) in spring 2016, 3 credit hours, 14 graduate students
- **Research Proposal Development (PLPT 892)** 1 credit hour (100% credit), taught fall 2016 only to my own first-year graduate students (1 in 2016)
- **Field Disease Tour (PLPT 891)** co-instructor (50% credit) in fall 2016, 1 credit hour, 14 graduate students

2015 CASNR Calculated FTE: 5.63% Instructional and 2.86% Advising

- **Developed new course** “Disease Dynamics and Evolution”, which targets upper level undergraduate students in microbiology and graduate students
- **Soil Biocontrol and Pathogens Journal Club** (non-credit) taught fall 2015 with total of 22 people that were signed up for our group and 10-15 attendees average
- **Research Proposal Development (PLPT 892)** 1 credit hour, taught fall 2015 and only to my own first-year graduate students (1 in 2015)
- **Independent Research (PLPT 892)** taught fall 2015 for student on rotation (1) at 2 credit hours

Activities in Scholarship of Teaching and Learning (SOTL)

- ARISE Learning by Design Workshop (taught by Sydney Brown), Fall 2017
- Peer Instruction Workshop (taught by Rob Erdmann & Marilyne Stains), Spring 2018
- Peer Review of Teaching Workshop (taught by Jody Kellas), Academic year 2015–2016

Benchmark Portfolio:

Everhart, S.E., "PLPT 496/892: Disease Dynamics & Evolution—A Peer Review of Teaching Project Benchmark Portfolio" (2016). UNL Faculty Course Portfolios. 20. <http://digitalcommons.unl.edu/prtunl/20>

Mentoring

Current lab members: Margarita Marroquin-Guzman (postdoc), Nikita Gambhir (Ph.D. student), Edgar Nieto Lopez (Ph.D. student), Karen Ferreira Da Silva (Ph.D. student), Rachel Persson (undergraduate UCARE student), Rebecca Higgins (research technologist)

Postdoctoral Scholars:

1. Margarita Marroquin, meta-barcoding soil microbial communities, June 2017–**present**
2. Zhian Kamvar, population genetics and genomics, January 2017–2018
3. Thomas Miorini, phenotyping *S. sclerotiorum*, March 2016–April 2017; January 2018 – 2018
4. B. Sajeewa Amaradasa, sub-lethal fungicide analysis of *S. sclerotiorum*, Aug. 2014 – Jul. 2016

Graduate Students:

5. Karen Ferreira Da Silva (Ph.D.), co-advised with Joe Louis on various projects, June 2018–**present**
6. Callie Braley (D.P.H.), co-advised with Loren Giesler on *C. sojina* fungicide sensitivity, Summer 2018
7. Gulcin Ercan (M.S.), co-advised w/ Adesemoye (chair) on entomopathogenic fungi, June 2017–**present**
8. Edgar Nieto Lopez (Ph.D.), resistance and population genetics of *S. sclerotiorum* from soybean and dry bean in the U.S. and Mexico, Aug. 2016—**present**
9. Nikita Gambhir (Ph.D.), resistance and population genetics in *R. solani*, and genomic analysis of *S. sclerotiorum* after sub-lethal fungicide exposure, Aug 2015 – **present**
10. Bridget Tripp (rotation), Complex Biosystems rotation student, conducted short project on reference-guided genome alignment for variant and marker development, Fall 2015

Undergraduate Research Assistants and Honors Thesis Students:

11. Rachel Persson, UCARE recipient and general laboratory worker, May 2018–**present**
12. Isabel Chavez, general laboratory work, November 2017–May 2018
13. Audrey Vega, general laboratory work, November 2017–May 2018
14. Alex Johnson, general laboratory and molecular research, July 2017–January 2018
15. Anthony Pannullo, fungicide sensitivity of *S. sclerotiorum* and Honors Thesis Research supported by IANR Undergraduate Research Award, May 2015–August 2017
Anthony is now a graduate student in the Department of Microbiology at the University of Iowa
16. Josh Hanson, culturing of *S. sclerotiorum*, Oct. 2014 – December 2017
17. Morgan Thompsen, general laboratory work, May 2016 – Dec. 2016.
18. Sarah Campbell, culturing and genotyping of *S. sclerotiorum*, Oct. 2014 – May 2016.
Sarah is now a graduate student in the Department of Plant Pathology at the University of Georgia
19. Flavio Nunes da Silva, isolation of *R. solani*, May-July, 2015 (10-week internship)

Research Technologists:

20. Rebecca Higgins, half-time appointment for *Sclerotinia* bean line screening, October 2018–**present**

Departmental Staff Hired and Supervised:

21. Jimin Kamvar, Digital Communications Liaison, October 2017–December 2017

FUNDING:**\$2.38 Million awarded in past 3 years (since Jan. 1, 2015)****Research Grants and Contracts:****\$1,920,249 total**

1. Proctor, C., **S.E. Everhart**, (and 9 others at 3 other institutions), “Optimizing cropping systems for resilience to stress: The role of maturity group selection and cover crops on yield, weeds, insects, and microbes”, USDA-NIFA Foundational on Pests and Beneficial Species, **\$461,187**, (2017 to 2020).
\$24,212 to SEE
2. Bond, J., **S.E. Everhart**, (and 13 others at 10 institutions), “Seedling diseases: Identification, management and education”, N. Central Soybean Research Program, **\$878,940**, (2015 to 2018). + *1-yr renewal for 2019*
\$55,500 to SEE
3. Kabbage, M., **S.E. Everhart**, (and 4 others at 3 institutions), “Biology and Control of Sclerotinia Stem Rot of Soybean”, N. Central Soybean Res. Program, **\$240,000**, (2015 to 2018). **\$75,000 to SEE**
4. **Everhart, S.E.**, and A. Adesemoye, “Fungicide resistance in *Rhizoctonia solani* and implications for soybean fields in Nebraska”, NE Soybean Board, **\$121,961**, (2015 to 2018). **\$102,804 to SEE**
4. Steadman, J., and **S.E. Everhart**, “Improved white mold resistance in dry and snap beans through multi-site screening and pathogen characterization throughout major production areas”, USDA-ARS National Sclerotinia Initiative, **\$192,536**, (2016 to 2018). + *1-yr renewal for 2019* **\$68,327 to SEE**
- Adams, G., and **S.E. Everhart**, “Population genetic analysis of the fungal pathogen *Gemmatomyces piceae* to determine native (sexual), introduced (clonal), or invasive (mixed) reproduction on spruce in Alaska”, USDA Forest Service, **\$10,000**, (2017 to 2018). **\$9,000 to SEE**
5. **Everhart, S.E.**, “Impact of sub-lethal fungicides on genome evolution: A potential new mechanism of resistance emergence in fungi”, UNL Layman Award, **\$10,000**, (2015 to 2016). **\$10,000 to SEE**
6. **Everhart, S.E.**, “*In vitro* fungicide testing of SDS pathogen, *Fusarium virguliforme* (current name *Neocosmopora virguliforme*)”, Gowan Company, **\$5,625**, (2016 to 2017). **\$5,625 to SEE**

Teaching and Fellowship Grants:**\$332,887 total**

7. Keshwani, J., **S.E. Everhart**, (and 3 others), “Cultivating ACCESS: Agriculture Career Communities to Empower Students in STEM”, USDA-NIFA WAMS: Women and Minorities in STEM Fields Program, **\$94,387**, (2017 to 2020). **\$5,000 to SEE**
8. Hein, G., **Everhart, S.E.**, (and 6 others), “Bridging the Gap: Educating multidisciplinary professionals to steward pest management technologies for sustainable agriculture”, USDA-NIFA National Needs Fellowship Program, **\$238,500**, (2016 to 2019). **\$0 to SEE**

Fellowship Support (with research proposals):**\$127,120 total**

9. Nieto-Lopez, E. (Graduate Student), and **S. Everhart (Advisor)**, Fellowship, “Fungicide Sensitivity and Population Structure of *Sclerotinia sclerotiorum* from Soybean in the North Central United States and Dry Bean in Mexico”, CONNACYT, Mexican Government, **\$124,620**, Awarded. (sub: April 2016, start: August, 2016, end: August, 2020) **\$0 to SEE**
10. Pannullo, A. (Undergraduate Student), and **S. Everhart (Advisor)**, Grant, “Population Genetic Diversity of *Sclerotinia sclerotiorum*, Causal Agent of White Mold Disease of Dry Bean, and Implications for Fungicide Resistance / Disease Management”, IANR Undergraduate Research Award, **\$2,500**, Awarded (sub: May 2016, start: July, 2016, end: May 2017) **\$2500 to SEE**

Travel Support:**\$1,500 total**

11. **Everhart, S.**, IC-1152, Grant, "2015 APS Annual Meeting", Internal, **\$500**, Awarded.
12. **Everhart, S.**, IC-1152, Grant, "2016 APS Annual Meeting", Internal, **\$500**, Awarded.
13. **Everhart, S.**, Award, “2016 Schroth Faces of the Future Award”, APS, **\$500**, Awarded